

## **JPred: a consensus secondary structure prediction server**

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### **Abstract**

**Summary:** An interactive protein secondary structure prediction Internet server is presented. The server allows a single sequence or multiple alignment to be submitted, and returns predictions from six secondary structure prediction algorithms that exploit evolutionary information from multiple sequences. A consensus prediction is also returned which improves the average Q<sub>3</sub> accuracy of prediction by 1% to 72.9%. The server simplifies the use of current prediction algorithms and allows conservation patterns important to structure and function to be identified.

**Availability:** <http://barton.ebi.ac.uk/servers/jpred.html>

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When predicting the secondary structure of a protein 'blind', without knowledge of the answer, it is useful to exploit the features of all available prediction algorithms rather than rely on one. Combination of methods has been applied successfully in a number of accurate predictions of protein secondary structure (e.g. see Edwards and Perkins, 1996; Crawford *et al.*, 1994; Russell *et al.*, 1992; Livingston and Barton, 1994, 1996; Russell and Barton, 1993). Unfortunately, combining prediction methods on a large scale is complicated by the fact that prediction programs have very different input requirements and output formats. In order to perform a recent large-scale comparative analysis of secondary structure prediction algorithms (Cuff and Barton, 1998), we developed flexible software to standardise the input and output requirements of six prediction algorithms. In this paper we describe a development of this work to provide the fully automatic JPred WWW server for multiple secondary structure prediction.

The server accepts two input types, a family of aligned protein sequences or a single protein sequence. If a single sequence is submitted, an automatic process creates a multiple sequence alignment, prior to prediction (Cuff and Barton, 1998).

Six different prediction methods (DSC (King and Sternberg, 1996), PHD (Rost and Sander, 1993), NNSSP (Sala-

mov and Solovyev, 1995), PREDATOR (Frishman and Argos, 1997), ZPRED (Zvelebil *et al.*, 1987) and MULPRED (Barton, 1988, unpublished)) are then run, and the results from each method are combined into a simple file format.

The NNSSP, DSC, PREDATOR, MULPRED, ZPRED and PHD methods were chosen as representatives of current state-of-the-art secondary structure prediction methods that exploit the evolutionary information from multiple sequences. Each derives its prediction using a different heuristic, based upon nearest neighbours (NNSSP), jury decision neural networks (PHD), linear discrimination (DSC), consensus single sequence method combination (MULPRED), hydrogen bonding propensities (PREDATOR), or conservation number weighted prediction (ZPRED).

The predictions and corresponding sequence alignment are rendered in coloured HTML, Java (Clamp *et al.*, 1998) and Postscript. The predictions are coloured and aligned with their corresponding family of sequences. Physico-chemical properties, solvent accessibility, prediction reliability and conservation number values (Zvelebil *et al.*, 1987) for each amino acid are included in the output. The original ASCII text data from each of the prediction methods can also be downloaded. For example, BLAST results (Altschul *et al.*, 1990), MSF and HSSP (Sander and Schneider, 1991) format alignments, pair comparison files and so on.

The text based MULPRED output is of particular interest as it is a combination of different single sequence prediction methods (Garnier *et al.*, 1978; Chou and Fasman, 1974; Lim, 1974; Rose, 1978; Wilmot and Thornton, 1988). While the automatic consensus within the MULPRED program is not as accurate as current methods (Cuff and Barton, 1998), the profile based output, when combined with the modern algorithms is helpful for human interpretation of the prediction.

A consensus prediction based upon a simple majority wins combination of NNSSP, DSC, PREDATOR and PHD is provided by the JPred server. If there is a tie, the prediction from PHD is used. In our independent test this approach gave the

highest accuracy compared to all other combinations (Cuff and Barton, 1998).

The consensus prediction achieved an average  $Q_3$  score of 72.9%, where  $Q_3$  is the percentage of residues predicted correctly for the three conformational states, strand helix and loop. This result is 1% better than PHD (71.9%) for the same data. The segment overlap score (Rost *et al.*, 1994) for the consensus method improves by 0.1%, to 75.4%. These results were obtained on a non-redundant set of 396 protein domain sequences that did not contain sequences similar to the proteins used to train the methods. Cuff and Barton (1998) includes a full analysis and description of the data sets, similarity cutoffs, accuracies and methods used for this test.

As well as providing a more accurate consensus prediction, the JPred server also permits the different prediction methods to be viewed concurrently with the alignment. This allows for easy interpretation and analysis of the prediction and multiple sequence alignment. Interactive analysis and re-alignment can also be carried out with the Java viewer and editor (Clamp *et al.*, 1998), where one may interactively change the colouring within the alignment to highlight important residues and conserved features.

In summary, JPred provides an automatic and simple to use tool to assist in accurate secondary structure prediction.

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## References

- Altschul,S.F., Gish,W., Miller,W., Myers,E.W. and Lipman,D.J. (1990) Basic local alignment search tool. *J. Mol. Biol.*, **215**, 403–410.
- Chou,P.Y. and Fasman,G.D. (1974) Conformational parameters for amino acids in helical, beta-sheet, and random coil regions calculated from proteins. *Biochemistry*, **13**, 211–222.
- Clamp,M.E., Cuff,J.A. and Barton,G.J. (1998) Jalview — a java multiple sequence alignment viewer and editor. <http://barton.ebi.ac.uk/>.
- Crawford,I.P., Niermann,T. and Kirchner,K. (1987) Prediction of secondary structure by evolutionary comparison: application to the alpha subunit of tryptophan synthase. *Proteins*, **2**, 118–129.
- Cuff,J.A. and Barton,G.J. (1998) Evaluation and improvement of multiple sequence methods for protein secondary structure prediction. *Proteins*, (submitted).
- Edwards,Y.J.K. and Perkins,S.J. (1996) Assessment of protein fold predictions from sequence information: the predicted alpha/beta doubly wound fold of the von Willebrand factor type A domain is similar to its crystal structure. *J. Mol. Biol.*, **260**, 277–285.
- Frishman,D. and Argos,P. (1997) Seventy-five percent accuracy in protein secondary structure prediction. *Proteins*, **27**, 329–335.
- Garnier,J., Osguthorpe,D.J. and Robson,B. (1978) Analysis and implications of simple methods for predicting the secondary structure of globular proteins. *J. Mol. Biol.*, **120**, 97–120.
- King,R.D. and Sternberg,M.J.E. (1996) Identification and application of the concepts important for accurate and reliable protein secondary structure prediction. *Protein Sci.*, **5**, 2298–2310.
- Lim,V.I. (1974) Algorithms for prediction of alpha helices and beta structural regions in globular proteins. *J. Mol. Biol.*, **88**, 873–894.
- Livingstone,C.D. and Barton,G.J. (1994) Secondary structure prediction from multiple sequence data: blood clotting factor xiii and yersinia protein tyrosine phosphatase. *Int. J. Pept. Prot. Res.*, **44**, 239–244.
- Livingstone,C.D. and Barton,G.J. (1996) Identification of functional residues and secondary structure from protein multiple sequence alignment. *Meth. Enz.*, **266**, 497–512.
- Rose,G.D. (1978) Prediction of chain turns in globular proteins on a hydrophobic basis. *Nature*, **272**, 586–591.
- Rost,B. and Sander,C. (1993) Prediction of protein secondary structure at better than 70% accuracy. *J. Mol. Biol.*, **232**, 584–599.
- Rost,B., Sander,C. and Schneider,R. (1994) Redefining the goals of protein secondary structure prediction. *J. Mol. Biol.*, **235**, 13–26.
- Russell,R.B. and Barton,G.J. (1993) An SH2-SH3 domain hybrid. *Nature*, **364**, 765.
- Russell,R.B., Breed,J. and Barton,G.J. (1992) Conservation analysis and structure prediction of the SH2 family of phosphotyrosine binding domains. *FEBS Letters*, **304**, 15–20.
- Salamov,A.A. and Solovyev,V.V. (1995) Prediction of protein secondary structure by combining nearest-neighbor algorithms and multiple sequence alignments. *J. Mol. Biol.*, **247**, 11–15.
- Sander,C. and Schneider,R. (1991) Database of homology-derived protein structures and the structural meaning of sequence alignment. *Proteins*, **9**, 56–68.
- Wilmot,A.C.M. and Thornton,J.M. (1988) Analysis and prediction of the different types of beta-turn in proteins. *J. Mol. Biol.*, **203**, 221–232.
- Zvelebil,M., Barton,G., Taylor,W. and Sternberg,M. (1987) Prediction of protein secondary structure and active sites using the alignment of homologous sequences. *J. Mol. Biol.*, **195**, 957–961.